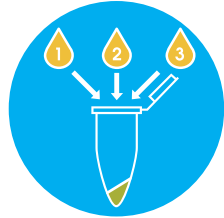


Viogene® Mini Plus™ Plasmid DNA Extraction System

Protocol for Vacuum Method

Operation Chart



Step 1 Step 2 Step 3
Add MX1, MX2, and MX3 buffer sequentially.



13,000rpm
5-10 min



Step 4
supernatant



Step 5
Vacuum



Step 6
Vacuum
add WN buffer



Step 7
Vacuum
add WS buffer



Step 8

13,000rpm
3 min



Step 9
add EB buffer

13,000rpm
1-2 min



Step 10

Cat.No:GF2001/GF2002

Step 1 : Add 200 μ l of **MX1 buffer** (RNase A added) to resuspend the cell Pellet vortexing or pipetting.

Step 2 : Add 250 μ l of **MX2 buffer** and gently mix (invert the tube 4-6 times) to lyse the cells until the lysate becomes clear. Incubate at room temperature for 1-5 minutes.

Step 3 : Add 350 μ l of **MX3 buffer** to neutralize the lysate, then immediately and gently mix the solution. A white precipitate should be formed.

Step 4 : Centrifuge for 5-10 minutes; meanwhile insert the tip of a Mini Plus Column into the luer-lock of a vacuum manifold.

Step 5 : Transfer the supernatant carefully into the column. Apply vacuum to draw all the liquid.

Step 6 : Wash the column once with 500 μ l **WN buffer** (ethanol added) by re-applying vacuum to draw all the liquid

Step 7 : Wash the column once with 700 μ l **WS buffer** (ethanol added) by re-applying vacuum to draw all the liquid.

Step 8 : Place the column onto a collection tube. Centrifuge the column at full speed for 3 minutes to remove ethanol residues.

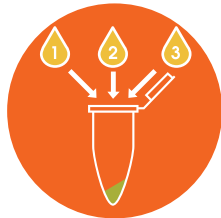
Step 9 : Place the column onto a new 1.5ml centrifuge tube. Add 50 μ l of **Elution buffer** (provided) at the center of the membrane. Stand the column for 1-2 minutes, and then centrifuge for 1-2 minutes to elute DNA.

Step 10 : Store plasmid DNA at 4°C or -20°C .

Viogene® Mini Plus™ Plasmid DNA Extraction System

Protocol for Centrifuge Method

Operation Chart



Step 1 Step 2 Step 3
Add MX1, MX2, and MX3 buffer sequentially.
cell peller

13,000rpm
5-10 min



Step 4
supernatant



Step 5



9,000rpm
30-60 sec



Step 6
add **WN** buffer

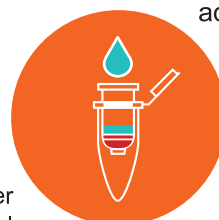
9,000rpm
30-60 sec



Step 7
add **WS** buffer

9,000rpm
30-60 sec

Centrifuge another
3 min at full speed
See Step 7.



Step 8
add **EB** buffer

13,000rpm
1-2 min



Step 9

Cat.No:GF2001/GF2002

Step 1 : Add 200 μ l of MX1 buffer (RNase A added) to resuspend the cell Pellet by vortexing or pipetting.

Step 2 : Add 250 μ l MX2 buffer and gently mix (invert the tube 4-6 times) to lyse the cells until the lysate becomes clear. Incubate at room temperature for 1-5 minutes.

Step 3 : Add 350 μ l of MX3 buffer to neutralize the lysate, then immediately and gently mix the solution. A white precipitate should be formed.

Step 4 : Centrifuge for 5-10 minutes, meanwhile place a Mini Plus Column onto a collection tube

Step 5 : Transfer the supernatant carefully into the column. Centrifuge for 30-60 seconds. Discard the flow-through.

Step 6 : Wash the column once with 500 μ l WN buffer (ethanol added) by centrifuge for 30-60 seconds. Discard the flow-through.

Step 7 : Wash the column once with 700 μ l WS buffer (ethanol added) by centrifuge for 30-60 seconds. Discard the flow-through. Centrifuge the column at full speed for another 3 minutes to remove ethanol residues.

Step 8 : Place the column onto a new 1.5 ml centrifuge tube. Add 50 μ l of Elution buffer (provided) at the center of the membrane. Stand the column for 1-2 minutes, and then centrifuge for 1-2 minutes to elute DNA.

Step 9 : Store plasmid DNA at 4°C or -20°C.